

Functional B-1 progenitor cells are present in the hematopoietic stem cell-deficient embryo and depend on *Cbfb* for their development

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The fetal liver is a major hematopoietic site containing progenitor cells that give rise to nearly all blood cells, including B-1 cells. Because the fetal liver is not a *de novo* site of hematopoietic stem cell (HSC) or progenitor-cell emergence, it must be seeded by yolk sac (YS)-derived erythromyeloid progenitors at embryonic day (E) 8.5–E10 and aorta-gonado-mesonephros (AGM)-derived HSCs at E10.5–E11.5. Although the B-1 progenitor cell pool in the fetal liver is considered to be of HSC origin, we have previously proposed that YS-derived B-1 progenitors may also contribute to this pool. Until now, it has been impossible to determine whether HSC-independent B-1 progenitor cells exist in the fetal liver. Here, we demonstrate the presence of transplantable fetal-liver B-1 and marginal zone B progenitor cells in genetically engineered HSC-deficient embryos. HSC-deficient YS and AGM tissues produce B-1 progenitors *in vitro* and thus may serve as sites of origin for the B-1 progenitors that seed the fetal liver. Furthermore, we have found that core-binding factor beta (*Cbfb*) expression is required for fetal-liver B-1 progenitor cell maturation and expansion. Our data provide, to our knowledge, the first evidence for the presence of B-1 progenitor cells in the fetal liver that arise independently of HSCs and implicate *Cbfb* as a critical molecule in the development of this lineage.

TEK | hemogenic endothelial cell | immune layered model

The immune layered model proposed by Herzenberg and Herzenberg in 1989 (1) predicted B-1 and B-2 lineage production through three waves of B lymphopoiesis arising from different precursors during development (2): According to this model, the first wave gives rise exclusively to innate immune B cells in early embryonic life and may be derived from progenitor cells independent of hematopoietic stem cells (HSCs), challenging the stem-cell theory that all blood cells are products of HSCs. The second and third waves are comprised of HSCs and HSC-derived progenitors in the fetal liver, neonatal bone marrow (BM) (the second wave), and adult BM (the third wave), respectively. Importantly, AA4.1⁺CD19⁺B220^{lo-neg} B-1 specific progenitors have been identified in the second wave (3). The second wave produces more B-1 cells than B-2 cells whereas the third wave displays an opposite skewing of B-cell differentiation (4–7). In fact, the B-1 cell-producing abilities of HSCs and common lymphoid progenitor cells decline with advancing age (6), and, in particular, CD5⁺B-1a cells are not produced by adult HSCs when examined by single HSC transplantation assay (7). Although the second and third waves have been examined in detail, it is unclear whether the first wave exists and contributes to innate immunity in postnatal life and whether the B-1 progenitor cells in wave 2 in the fetal liver are all HSC-derived or contain derivatives of the wave 1 HSC-independent embryonic progenitor cells.

Murine B-1 cells are innate immune cells (distinguished from conventional B-2 cells by specific surface markers such as

IgM^{hi}IgD^{lo}CD11b⁺), residing in the peritoneal and pleural cavities. These cells produce stereotypic natural antibodies in a T cell-independent manner and execute important roles in the first line of defense against microbial infection (8, 9). B-1 cells are segregated into CD5⁺B-1a and CD5[−]B-1b cells. Marginal zone (MZ) B cells, named after the restricted localization of these cells in the splenic marginal zone, are usually categorized as BM HSC-derived B-2 cells but share similar functions with B-1 cells, such as rapid production of IgM antibodies against bacterial pathogens in a T cell-independent manner. There is evidence that a portion of MZ B cells is also of embryonic or fetal origin (10–12).

We have recently reported that yolk sac (YS) and para-aortic-splanchnopleura (P-Sp) hemogenic endothelial cells (HECs) harvested before the first emergence of HSC give rise to transplantable, functional B-1a, B-1b, and MZ B cells *in vitro* and thus have provided supportive evidence for the first wave of B cells (13). However, because we isolated and cultured YS/P-Sp cells *in vitro* to allow them to differentiate into B-1 progenitor cells, whether YS/P-Sp-derived B progenitor cells seed the fetal liver *in vivo* and contribute to the B-1 progenitor cell pool or mature B-1 or MZ B cells in postnatal life has never been established. In other words, to address the question whether the first wave of B lymphopoiesis is present *in vivo* or not, we have to confirm the existence of HSC-independent B-1 progenitor cells in the fetal liver.

The fetal liver is an organ dependent upon hematopoietic stem/progenitor cell seeding from different hematopoietic tissues.

Significance

All lymphoid cells are considered to be products of hematopoietic stem cells (HSCs); however, it has been suggested, but not proven, that innate immune B-1 progenitor cells develop independently of HSCs in the fetal liver. B-1 cells, especially B-1a cells, are not replaced by adult bone marrow transplantation. Thus, it is critical to understand the origin and mechanisms required to sustain these cells *in vivo* because B-1 cells play important roles in the first line of defense against microbial infection and in preventing organ damage in autoimmune patients and infections in some patients after bone-marrow transplantation. We demonstrate that B-1 progenitor cells can develop independently of HSCs in the fetal liver and that their development relies critically on the expression of core-binding factor beta.

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It is an established concept that erythro-myeloid progenitors (EMPs) derived from embryonic day (E) 8.5–E10 YSs seed the fetal liver to support homeostatic hematopoiesis in the embryo whereas HSCs that emerge in the aorta-gonado-mesonephros (AGM) region seed the fetal liver at E11 and provide hematopoietic support later in development (14, 15). However, it is unknown whether the YS/P-Sp HEC-derived B-1 lymphoid progenitors seed the fetal liver. Because the B-1 progenitor cell pool in the fetal liver is considered to be an HSC derivative and because HSCs exist in the fetal liver concomitant with B-1 progenitor cells, it has been impossible to prove the existence of HSC-independent B lymphopoiesis in the fetal liver. To specifically address this question, we have used a unique mouse model devoid of HSC but known to possess some uncharacterized fetal-liver B cells (16, 17).

Core-binding factor beta (CBF β) is the common non-DNA-binding subunit of the Cbf family of heterodimeric transcription factors. By associating with CBF α subunits, CBF β increases the affinity of CBF α DNA-binding. Runt related transcription factor 1 (*Runx1*) also called core-binding factor alpha 2 (*Cbfa2*)^{-/-} embryos die at E12 and display severe deficiencies in definitive EMP, lymphoid cells, and HSCs (18). CBF β is required for Runx1 function, and loss of CBF β in embryos leads to embryonic lethality at E11–E13 similar to *Runx1*^{-/-} embryos with a severe defect in definitive hematopoiesis and HSCs (17, 19). However, when *Cbfb* expression was restored in *Cbfb*^{-/-} embryos under transgenic control of the endothelial promoter endothelial-specific receptor tyrosine kinase (*Tek*) (*Cbfb*^{-/-}:*Tek-GFP/Cbfb*), EMP formation and rare B and T cells were observed (16). In these rescued embryos, lymphoid progenitors (Lin⁻c-Kit⁺Flt3⁺) were detectable in the E14 fetal liver and produced B or T cells in OP9 or OP9-DL1 in vitro cocultures, respectively. However, E14 *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells failed to reconstitute recipient peripheral blood (PB) and BM upon transplantation, indicating that *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos contain lymphoid progenitors without transplantable HSCs. It has been previously reported that lymphoid progenitor cells are present in the YS and P-Sp before HSC emergence at an early stage (E8.25–E9.5) of embryo development (13, 20–23). Furthermore, the presence of multipotent progenitor cells with myeloid, B-cell, and T-cell lineage potentials without long-term HSC activity also has been reported (24–26). Whether the B and T progenitor cells emerge from a common HEC or are independently generated has not been fully resolved. Nonetheless, the fact that lymphoid progenitor cells are present prior to HSC emergence suggests that the initial phases of developmental hematopoiesis may not follow the same paradigm as adult BM HSC development where all of the lymphoid progenitor cells are HSC-derived (15, 27). Indeed, transplantable HSCs were rescued when *Cbfb* was expressed under transgenic control of the *Ly6a* promoter in the *Cbfb*^{-/-} embryos (*Cbfb*^{-/-}:*Ly6a-GFP/Cbfb*), suggesting that *Cbfb* is required in the *Ly6a*⁺ HECs for producing transplantable HSCs that repopulate myeloid and lymphoid lineages (16). Based on these findings and our previous reports that B-1 (but not B-2) and T lymphoid progenitor cells are present in the embryo before HSC emergence (13, 20), we hypothesized that the unspecified B cells detected in the *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos that do not harbor transplantable HSCs are HSC-independent B-1 progenitor cells that constitute the first wave of B lymphopoiesis.

In the present study, we confirm the existence of fetal-liver AA4.1⁺CD19⁺B220⁻ B-1 progenitor cells in the absence of transplantable HSCs in vivo. YS and AGM cells from *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos produce B-1 progenitor cells in vitro. Our results clearly demonstrate the presence of B-1 and MZ B progenitor cells in the fetal liver independent of HSCs and suggest that these progenitor cells are likely to be derived in part from YS/P-Sp sites. However, these B-1 progenitor cells do not display persistent expression of *GFP/Cbfb* because *TEK* is not

expressed once HECs become hematopoietic cells. Therefore, this *Cbfb* deficiency in B-1 progenitor population results in impaired maturation and expansion of these cells. Consistently, we show that overexpression of *Cbfb* in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells via a retrovirus promotes engraftment of B-1 and MZ B cells in recipient immunodeficient mice. Thus, our findings demonstrate that persistent *Cbfb* expression is required for the maturation and expansion of B-1 progenitor cells during in vivo development.

Results

B-1 Progenitor Cells Exist in the Fetal Liver in the Absence of HSCs.

First, we examined whether AA4.1⁺CD19⁺B220^{lo-neg} B-1 progenitor cells exist in the fetal liver of *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos. As previously reported (16), *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos can survive until birth in the absence of HSCs. The fetal livers of *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos at E15.5 and E17.5 were of similar size to the fetal liver of wild-type (WT) littermate embryos. Total fetal-liver mononuclear cell (MNC) counts from E17.5 *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos were significantly lower than WT embryos (Fig. 1B). As shown in Fig. 1A, lin⁻AA4.1⁺CD19⁺B220^{lo-neg} B-1 progenitor cells were detected in the fetal livers of *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos from E15.5 until birth although the frequency and total number were significantly lower than WT values (Fig. 1A–D and M). Furthermore, lin⁻AA4.1⁺CD19⁺B220^{lo-neg} B-1 progenitor cells were also detected in the fetal spleen from E15.5 until birth (Fig. 1G–J and N). Few AA4.1⁺CD19⁺B220⁺ B cells were detected in the fetal liver and spleen of *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos whereas WT fetal liver contained more AA4.1⁺CD19⁺B220⁺ B cells than B-1 progenitor cells (Fig. 1A, E–G, K, and L). AA4.1⁺CD19⁺B220⁺ cells are overlapping with AA4.1⁺B220⁺CD43⁺ proB cells (5). Whereas adult BM AA4.1⁺CD19⁺B220⁺ cells give rise predominantly to B-2 cells, fetal-liver AA4.1⁺CD19⁺B220⁺ or AA4.1⁺B220⁺CD43⁺ cells are known to give rise mainly to B-1 cells upon transplantation (3, 5). Thus, the fetal-liver AA4.1⁺CD19⁺B220⁺ population contains more B-1 lineage than B-2 lineage cells and is considered to be differentiated from both HSCs (through AA4.1⁺CD19⁻B220⁺ phenotype) and B-1 progenitor cells as we have previously reported (13). Therefore, severe reduction of AA4.1⁺CD19⁺B220⁺ cells in the fetal liver (Fig. 1E and F) and spleen (Fig. 1K and L) is due to HSC deficiency and an impaired maturation of HSC-independent B-1 progenitor cells in the *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos. Nevertheless, it is of note that *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal spleen contains B progenitor cells ($2.0 \times 10^4 \pm 1.7 \times 10^4$ cells) at 44% of the number measured in WT fetal spleen ($4.7 \times 10^4 \pm 1.3 \times 10^4$ cells) (by calculating total B progenitor cells in E17.5 spleen) (Fig. 1N). This result suggests that at least 40% of B-1 progenitor cells in the fetal-neonatal spleen are HSC-independent, a point that contrasts with the common perception that all lymphoid cells are HSC-derived.

B-1 Progenitors in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* Fetal Liver Fail to Mature and Expand in Vitro.

Although we have confirmed for the first time, to our knowledge, that B-1 progenitors are present in the HSC-deficient fetal liver, we have also observed a possible maturation defect of B-1 progenitor cells in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos (Fig. 1). To examine the maturation capacity of B-1 progenitor cells in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal liver, AA4.1⁺CD19⁺B220^{lo-neg} B-1 progenitor cells from WT and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal liver were plated on OP9 stromal cells with added IL-7 and Flt3-ligand. WT AA4.1⁺CD19⁺B220^{lo-neg} cells proliferated, forming cobblestones beneath the stromal cells, and produced more than 10^6 nonadherent cells with CD19⁺B220⁺ (including IgM⁺) B cells in 7–10 d (Fig. 2A, Upper). In contrast, *Cbfb*^{-/-}:*Tek-GFP/Cbfb* AA4.1⁺CD19⁺B220^{lo-neg} cells did not proliferate as robustly although they produced cobble stones by

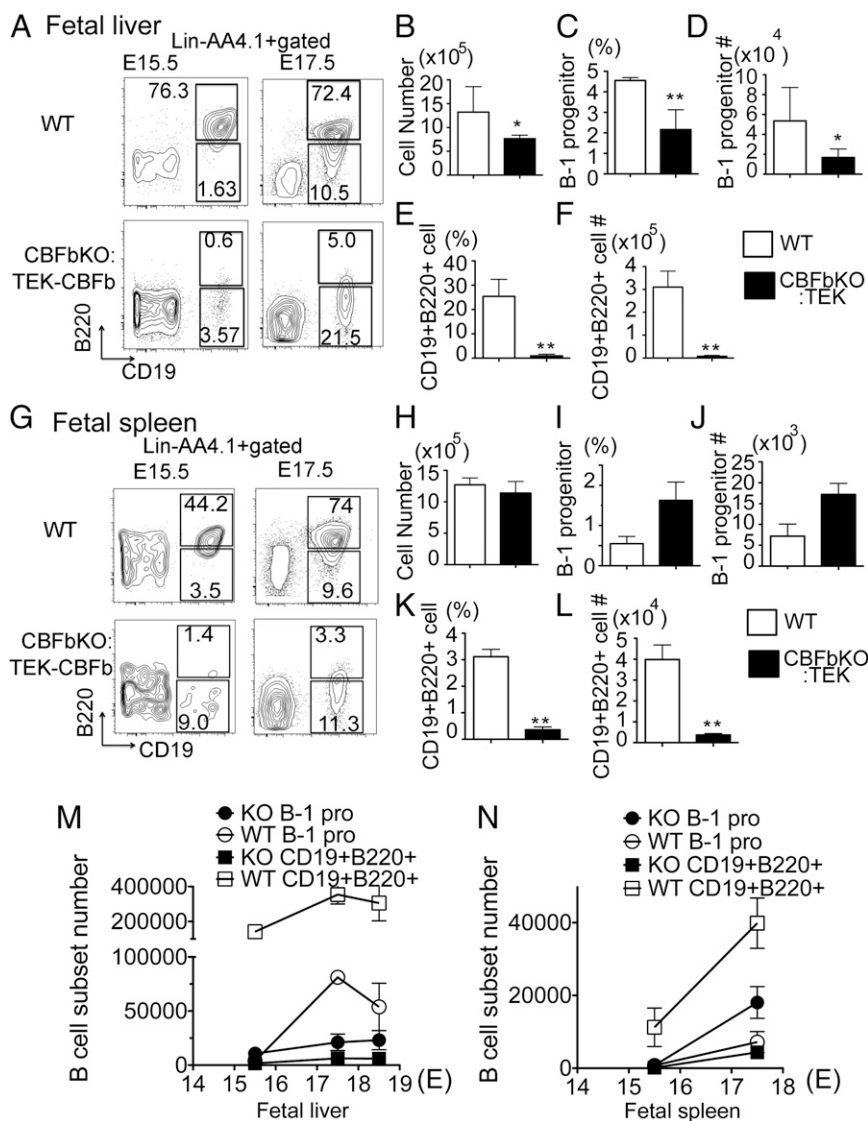


Fig. 1. B-1 progenitor cells exist in the *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal liver (FL) and spleen. AA4.1⁺CD19⁺B220^{lo-neg} B-1 progenitor phenotype was examined in the WT and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver and spleen cells at each day from E15.5 until birth (E18.5). The representative data for E15.5 and E17.5 are depicted. Lin⁻AA4.1⁺ gated FACS dot plots at E15.5 and E17.5 fetal liver (A) and fetal spleen (G) are depicted. Total MNCs of E17.5 FL (B) and spleen (H), percentage of B-1 progenitor cells in FL (C) and spleen (I), total cell number of B-1 progenitor cells in FL (D) and spleen (J), percentage of AA4.1⁺CD19⁺B220⁺ cells in FL (E) and spleen (K), and total cell number of AA4.1⁺CD19⁺B220⁺ cells in FL (F) and spleen (L) are shown. (B–F and H–L) Open bar, WT; filled bar, *Cbfb*^{-/-}:*Tek-GFP/Cbfb*. The cell numbers of B-1 progenitors and CD19⁺B220⁺ cells from WT and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* FL (M) and spleen (N) at each embryonic day are shown (*n* = 4 for each group; **P* < 0.05, ***P* < 0.01).

17 d with 10^6 nonadherent cells, including a few CD19⁺B220⁺ cells and many Mac1⁺ macrophages (Fig. 2A, Lower). Considering the fact that AA4.1⁺CD19⁺B220^{lo-neg} cells were originally reported as B cell-macrophage bipotent progenitors (28), these results indicated that loss of *Cbfb* may have contributed to the reduction of B lymphoid potential in this population. This is compatible with a previous report that the B lymphoid potential of lymphoid progenitor cells in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos was 30-fold less than in WT embryos (16). Thus, proliferation and B-1 cell differentiation capacity were significantly impaired in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* B-1 progenitor cells at least under the coculture conditions we used in vitro.

YS and AGM Can Produce B-1 Progenitor Cells in the Embryo Devoid of HSCs. The fetal liver is not a de novo site of hematopoietic cell emergence and must be seeded by various hematopoietic progenitor/stem cells derived from other hematopoietic tissues (15). Because HSCs are not produced in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos, we examined the possible source of B-1 progenitor cells that seed the fetal liver. We cocultured YS and P-Sp/AGM cells with OP9 stromal cells as previously reported (13). As expected, *Cbfb*^{-/-}:*Tek-GFP/Cbfb* YS and AGM cells produced AA4.1⁺CD19⁺B220⁺ B cells but at a lower frequency compared with WT YS/AGM (Fig. 2B). These cultured YS/P-Sp/AGM-derived

AA4.1⁺CD19⁺B220⁺ B progenitor cells were confirmed to become mature B-1 cells in vivo upon transplantation as previously reported (13). Although there is a study suggesting that placenta has the potential of producing B cells at a pre-HSC stage (29), we found maternal B cells in the placenta at this time point whereas YS and P-Sp derived B-1 cells were confirmed to be embryonic in origin (13). Thus, although we cannot exclude the possibility of the placenta as a production site for the first wave of B-1 progenitor cells in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos, our data indicate that YS and AGM produce B-1 progenitor cells that may seed the fetal liver in the *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos devoid of HSCs.

B-1 Progenitor Cells Can Mature into B-1 and MZ B Cells in Vivo but at Low Efficiency. Lin⁻AA4.1⁺CD19⁺B220^{lo-neg} B-1 progenitor cells were maintained until birth in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* mice (B progenitor cells were present at 40% of the number of WT mice in the spleen) (Fig. 1N), and mice died 1 d after birth as previously reported (16). Because mature B-1 cells are not detectable in the peritoneal cavity until 5 d after birth (30), we were unable to find mature peritoneal B-1 cells in the *Cbfb*^{-/-}:*Tek-GFP/Cbfb* live-born neonates. To examine whether the B-1 progenitor cells found in the fetal liver of *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos can mature into B-1 cells in vivo, we transplanted fetal-liver MNCs

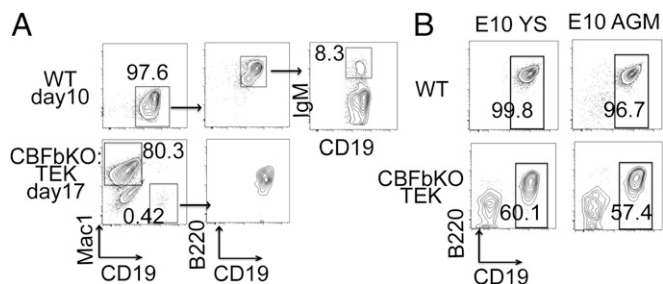


Fig. 2. (A) B-1 progenitor cells from *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal liver fail to expand in vitro. AA4.1⁺CD19⁺B220^{lo-neg} B-1 progenitor cells were sorted from E17.5 WT and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* FL and plated on OP9 stromal cells with IL-7 and Flt3-ligand. FACS dot plots of WT culture on day 10 (Upper) and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* culture on day 17 (Lower) are depicted. Representative FACS dot plots from three separate experiments are depicted. (B) YS and AGM cells from *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos can produce B progenitor cells in vitro. E10 YS and AGM from WT (Upper) and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* (Lower) embryos were plated on OP9 stromal cells with IL-7 and Flt3-ligand. Cells after 10-d coculture were analyzed for their surface markers by flow cytometry. Representative FACS dot plots from two separate experiments (each experiment was done in triplicate) are depicted.

from WT and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos into irradiated NOD/SCID/IL2γ^{-/-} (NSG) neonates. Although WT fetal-liver MNCs displayed multilineage repopulation in the PB of the recipient mice (Fig. 3A, Upper), no apparent engraftment was found in the PB of the recipient mice transplanted with *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver MNCs as previously reported (16) (Fig. 3A, Lower). However, in the host peritoneal cavity and spleen, B-1 and MZ B cells were reconstituted from donor *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver MNCs (Fig. 3B). No donor-derived B-2 cells or T cells were detected (Figs. 3B and 4E). These data indicated that HSC-deficient fetal-liver MNCs contain progenitor cells that can mature into only B-1 and MZ B cells in vivo upon transplantation.

Cbfb Is Required for Maturation and Expansion of B-1 and MZ B Cells.

Although transplantable B-1 and MZ B-cell progenitors are present in HSC-deficient fetal liver, the level of chimerism was significantly lower than donor contributions from WT embryos (WT 97.8 ± 1.3% vs. *Cbfb*^{-/-}:*Tek-GFP/Cbfb* 1.98 ± 0.3% in the peritoneal cavity and WT 92.8 ± 3.3% vs. *Cbfb*^{-/-}:*Tek-GFP/Cbfb* 3.4 ± 2.4% in the spleen). It has been reported that *Cbfb* is required in all blood-lineage cells for their development, but *TEK-GFP/Cbfb* expression was not sustained once *Tek-GFP/Cbfb*⁺ HECs become hematopoietic cells (16, 17). We confirmed that WT fetal-liver B-1 progenitor cells express *Cbfb* at a similar level to HSCs (Fig. 4A). However, B-1 progenitor cells from *Cbfb*^{-/-}:*Tek-GFP/Cbfb* do not express *TEK-GFP/Cbfb* (Fig. 4B), thus having lost *Cbfb* expression during differentiation from *TEK*⁺ HEC to B-1 progenitor cells (Fig. 4C). We have also recently determined that loss of *Runx1* or *Cbfb* in the B-cell lineage caused B-2 cell maturation defects (31). In this mouse model of *Cbfb* deficient in B-cell lineage, we also observed a threefold reduction in B-1 cell number compared with WT mice, indicating that *Cbfb* plays a critical role in B-1 cell development (Fig. S1) (31). Thus, we hypothesized that *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver B-1 progenitor cells would require ongoing *Cbfb* expression to mature into functional B-1 cells. To test the hypothesis, we overexpressed *Cbfb* in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver MNCs using a retrovirus vector that constitutively produced Cbfb (Fig. 4D and Fig. S24). Once retrovirally infected fetal-liver *Cbfb*^{-/-}:*Tek-GFP/Cbfb* MNCs were transplanted into irradiated NSG neonates, *Cbfb*^{-/-}:*Tek-GFP/Cbfb*-derived cells displayed dramatically improved engraftment capacity in up to 49% of donor cells

in the peritoneal cavity (Fig. 4E) and improved maturation into IgM⁺ cells in the spleen (Fig. S3), and displayed only B-1 cell reconstitution in the peritoneal cavity and MZ and B-1 cells in the spleen 12 wk after transplantation (Fig. 4E and F and Fig. S3). Nearly all (98%) of these *Cbfb*^{-/-}:*Tek-GFP/Cbfb*-derived cells expressed retroviral Cbfb-GFP (Fig. S2B) and displayed 15–25 times more *Cbfb* mRNA expression compared with WT peritoneal B-1 cells (Fig. S2C). In contrast, *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells infected with the empty vector control virus poorly engrafted (Fig. 4E, Middle). These results indicate that *Cbfb* overexpression rescues maturation and expansion of impaired B-1 progenitor cells from *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos. Furthermore, *Cbfb* expressing engrafted B-1 cells in the recipient mice were functional, displaying T cell-independent antiphosphorylcholine IgM secretion detected by enzyme-linked immunospot (Elispot) assay (Fig. 4G and H). Taken together, *Cbfb* reexpression restored engraftment and expansion/maintenance capacity of HSC-independent B-1 and MZ B cells from *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal liver in vivo, suggesting that *Cbfb* plays important roles in B-1 cell development and proliferation, similar to B-2 cell development (31). Of note, HSCs were not restored by retrovirus *Cbfb* overexpression in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells: no engraftment of any hematopoietic lineages other than B-1 or MZ B cells (Fig. 4E, Bottom, Fig. 4F, and Fig. S3), suggesting that the timing and dose of *Cbfb* expression may be differentially required for the development of each hematopoietic lineage.

Discussion

In the current study, using unique HSC-deficient embryos, we have for the first time, to our knowledge, proven that progenitor cells that give rise to only B-1 and MZ cells exist in the fetal liver in the absence of HSCs. Although we were not able to quantitatively estimate the percentage contribution of the first wave of B cells in postnatal life (because the *Cbfb*^{-/-}:*Tek-GFP/Cbfb* neonates die just after birth), it is probable that at least 40% of

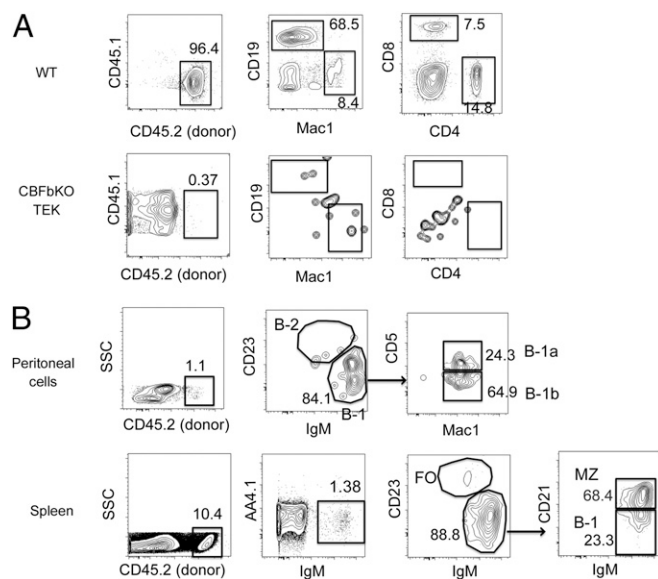


Fig. 3. *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells contain a few B-1 and MZ progenitor cells, but not HSCs. E15.5 FL MNCs from WT and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos were transplanted into sublethally irradiated NSG neonates. PB analysis 8 wk after injection is depicted (A). PB of recipient mice transplanted with WT (Upper) and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* (Lower) fetal-liver cells are depicted. (B) The peritoneal (Upper) and spleen (Lower) cells in the recipient mice transplanted with *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells are shown. FO, follicular B cell.

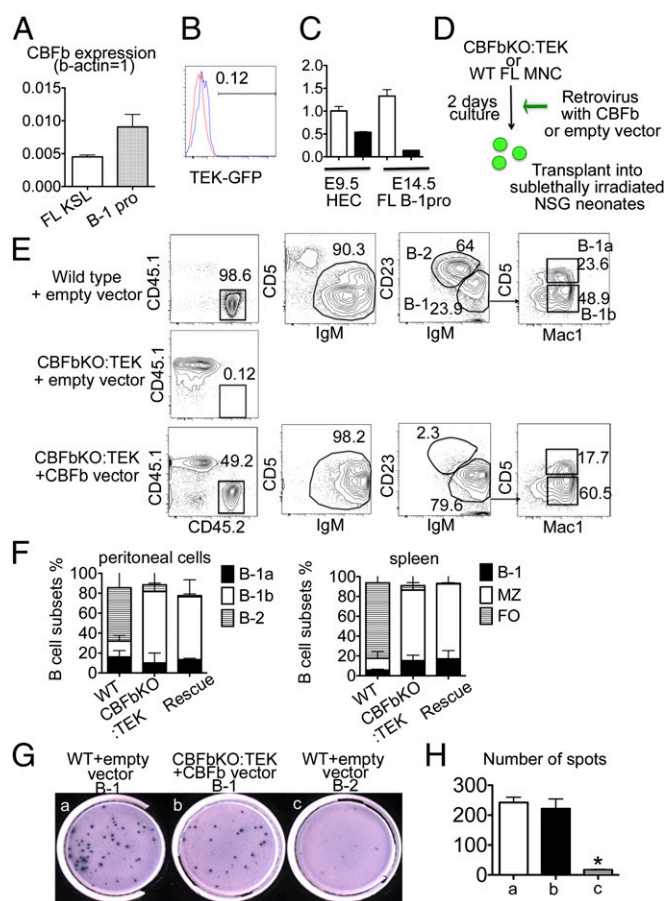


Fig. 4. Overexpression of *Cbfb* in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells restored B-1 and MZ cell engraftment. (A) *Cbfb* expression in WT fetal-liver HSCs and B-1 progenitors by quantitative PCR is shown. Internal control: β -actin = 1.0. (B) *Tek-GFP/Cbfb* expression within E15.5 fetal-liver AA4.1⁺ CD19⁺B220^{dim} B-1 progenitor cells in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos. Red line, WT; blue line, *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver B-1 progenitor cells. (C) Relative *Cbfb* expression in E9.5 WT CD31⁺Tie2⁺ HECs, E9.5 *Cbfb*^{-/-}:*Tek-GFP/Cbfb* HECs, E14.5 WT FL B-1 progenitors, and E14.5 *Cbfb*^{-/-}:*Tek-GFP/Cbfb* FL B-1 progenitors, compared with E9.5 WT HECs (=1). Open bar, WT; filled bar, *Cbfb*^{-/-}:*Tek-GFP/Cbfb*. (D) Experimental design for rescuing *Cbfb* expression in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells. (E) Peritoneal cells of recipient mice at 12 wk after transplantation with fetal-liver cells from WT (Top) or *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells expressing empty vector (Middle) or *Cbfb* (Bottom), respectively. (F) Donor-derived B-cell subsets in the recipient peritoneal cavity (Left) and spleen (Right) are depicted. Donor cells were WT fetal-liver MNC cells (left bars), *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells (center bars), and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells infected with *Cbfb* expressing retrovirus (rescue, right bars), in each panel. (G) IgM secretions from B-1 or B-2 cells stimulated with phosphorylcholine were detected by Elispot assay: (a) 1,000 B-1 cells from the recipient mice transplanted with WT fetal-liver cells with empty vector, (b) 650 B-1 cells derived from *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal-liver cells with *Cbfb* expression vector, and (c) 1,000 B-2 cells derived from WT fetal-liver cells with empty vector were initially plated. (H) The numbers of spots per 1,000 B-1 or B-2 cells derived from recipient mice in Elispot assay (G, a, b, and c) are depicted. There was no significant difference between B-1 cells from WT fetal liver with empty vector and from *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal liver with *Cbfb* expression vector. The number of IgM-secreting B-2 cells was significantly less than that of B-1 cells from WT or *Cbfb*^{-/-}:*Tek-GFP/Cbfb* with *Cbfb* expression vector (<0.05).

neonatal spleen B cells are derived from HSC-independent progenitors. Since fetal-liver cells do not produce any blood cells de novo but rely upon seeding from other sites, these B-1 progenitor cells must have been seeded from another site(s) of embryonic hematopoiesis, such as the yolk sac or embryo proper.

We also report that progenitor cells for MZ B cells developed in the absence of HSCs in the *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos. We previously identified YS/P-Sp-derived B-1 progenitor cells as giving rise to MZ B cells upon transplantation in vivo (13). The *Cbfb*^{-/-}:*Tek-GFP/Cbfb* mouse model was instrumental in again validating the HSC-independent origin of MZ B cells. Because MZ B cells are also produced by adult BM HSCs and splenic lin⁻CD19⁺B220^{lo-neg}CD43⁻ precursors (7, 32), we speculate that MZ B cells may comprise precursors from heterogeneous sites. Ito et al. have reported less frequent generation of MZ B than B-1 cells from YS lymphoid progenitors (33) but still showed some MZ B-cell potential of the YS precursors. Differences in their results from our present work may be the result of differences in coculture lines and choice of cytokines or cytokine concentrations. Ectopic expression of Lin28b has been reported to reprogram adult hematopoietic stem/progenitor cells into cells with the ability to give rise to fetal lymphopoiesis (34). In this report, Lin28-expressing cells differentiated into predominantly B-1a and MZ B cells in the recipient mice, in agreement with our results. Thus, accumulating evidence supports a paradigm that a part of MZ B cells are derived from embryonic precursors, similar to B-1 cells.

Another important finding from our current work is that *Cbfb* expression is required for B-1 cell development as well as B-2 cell development as previously reported (31). However, B-1 cell maturation is permissive in *Cbfb*^{-/-}:*Tek-GFP/Cbfb* fetal liver-derived B-1 progenitor cells, and the reduction of mature B-1a cells in *Cbfb*^{lox/lox}:*MB1-cre* mice is milder than the B-2 lineage. *Cbfb* may play different roles in the B-1 and the B-2 cell lineages, and further study would be required to fully explore the roles played in each lineage.

In sum, the presence of transplantable B-1 and MZ progenitor cells in the fetal liver devoid of HSCs confirms the first wave of B lymphopoiesis in the immune layered model predicted more than 20 y ago (1). Many questions remain to be addressed. How do B-1 and MZ B-cell progenitors migrate into the fetal liver, spleen, and BM? How and where do the B-1 progenitor cells mature? How much do they really contribute to innate immunity compared with fetal-liver HSC-derived B-1 and MZ B cells? Lineage-tracing models would be required to resolve these questions, and answering those questions would help to understand the roles of B-1 cells in autoimmunity (35, 36), prevention of atherosclerosis (37), and prevention of infections in blood stem cell transplanted patients (38).

Materials and Methods

Mice. *Cbfb*^{+/-}:*Tek-GFP/Cbfb* mice were generated by crossing *Cbfb*^{+/-} mice and *Tek-GFP/Cbfb* mice (kindly provided by Dr. Nancy Speck, University of Pennsylvania, Philadelphia). To obtain *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos, timed mating was performed by crossing *Cbfb*^{+/-}:*Tek-GFP/Cbfb* males with *Cbfb*^{+/-}:*Tek-GFP/Cbfb* or *Cbfb*^{+/-} females. Embryonic tail was used for genotyping.

Transplantation. NSG neonates were used as recipients. One- to 5-d-old neonates were sublethally irradiated (150 rad), and fetal-liver cells from WT and *Cbfb*^{-/-}:*Tek-GFP/Cbfb* embryos were injected into the peritoneal cavity of NSG neonates. Six to 12 wk after injection, PB, peritoneal, and spleen cells were analyzed donor cell types in the recipient mice.

OP9 Culture. An OP9 stromal cell line was maintained with α -MEM and 20% (vol/vol) FBS. YS, P-Sp/AGM, or fetal-liver cells were plated on six-well plates confluent with OP9 stromal cells in induction medium [α -MEM, 10% (vol/vol) FBS and 5×10^{-5} M β -mercaptoethanol] supplemented with 10 ng/mL IL-7 and 10 ng/mL Flit3-ligand (PEPROTECH) as previously reported (13). Floating cells in the culture medium were collected at optimal time points, and we analyzed their surface markers by flow cytometry.

Flow Cytometry. For analysis and sorting of B-1 progenitors, B-1, MZ B Cells, and each lineage of blood cells, the following antibodies were used at different fluorescent color combinations: anti-mouse AA4.1 (AA4.1), CD19 (1D3), B220 (RA3-6B2), IgM (II/41), CD21 (8D9), CD23 (B3B4), CD11b (M1/70),

CD5 (53-7.3), CD3e (145-2C11), Ter119 (TER-119), c-kit (2B8), Sca-1 (D7), and IL-7Ra (A7R34) (all purchased from eBioscience). A lineage mixture containing anti-Ter119, CD11b, and CD3 was used for lineage negative gating. Cells were analyzed on LSRII or sorted on FACS Aria (Becton Dickinson).

Quantitative PCR. Fetal-liver Lin⁻Sca-1⁺c-kit⁺ cells or AA4.1⁺CD19⁺B220^{lo-neg} B-1 progenitors were sorted, and RNA was extracted using RNasy (QIAGEN) following the manufacturer's instructions. cDNA was made using Super Script III fast strand Synthesis System (Invitrogen). Real-time PCR was performed using FastStart Universal SYBR Green Master (Rox) (Roche) by 7500 Real-Time PCR System (Life Technologies). The following sets of primers were used: *Cbfb*, forward, GCAAGTTCGAGAACGAGGAG; reverse, CCCGTGTACT-TAATCTCGCA; β -actin, forward, TCCTGTGGCATCCATGAAACT; reverse, GAAG-CACTTGCAGTGCACGAT.

Expression Vectors and Retrovirus Production. Full-length cDNA of mouse *Cbfb* was purchased from TrueClone (catalog no. MC204426) and amplified by PCR. cDNA of mouse *Cbfb* was inserted into an MSCV-IRES-GFP (MIG) plasmid. Retroviruses carrying empty vector (MIG-empty) or MIG-*Cbfb* were produced using the Phoenix cell line. Virus supernatants were concentrated by centrifuging at 19,000 \times g, 4 °C for 100 min. WT and *Cbfb*^{-/-}: *Tek-GFP*/*Cbfb* fetal-liver MNCs were plated on retronectin-coated 24-well plates in induction medium with 10 ng/mL stem cell factor, 10 ng/mL IL-7, and 10 ng/mL Flt3-ligand. Concentrated virus supernatant was added into each well, and spin infection was performed by centrifuging at 1,000 \times g, 32 °C for 100 min.

The next morning, the medium was half changed, and second-spin infection was done. Three hours after second infection, cells were harvested and used for the transplantation assay. Infectious efficiency was confirmed by detecting GFP⁺ cells by flow cytometry.

Elispot. Elispot was done as previously described (39). Donor-derived B-1 or B-2 cells in the peritoneal cavity of recipient mice were sorted on a FACS Aria (Becton Dickinson), suspended in Elispot medium (RPMI 1640 medium with 10% FBS and 5 \times 10⁻⁵ M β -mercaptoethanol), supplemented with 10 ng/mL IL-5, and cultured for 36–48 h in 96-well plates precoated with phosphor-ylcholine-conjugated keyhole limpet hemocyanin. Five hundred to 1,000 cells were distributed into an Elispot MultiScreen^{HTS} Filter Plate (Millipore) precoated with anti-mouse IgM and cultured in Elispot medium overnight. After washing, alkaline phosphatase-conjugated goat anti-mouse IgM (Southern Biotech) was applied to each well and incubated for 45 min. After washing, spots were visualized using BCIP/NBTplus substrate as described (Mabtech).

Statistics. All experiments were completed in triplicate. The Student t test was used for statistical analysis.

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